



Valorisation de la biodiversité microbienne pour les aliments fermentés : analyse multi-échelle du génotype au phénotype pour un choix rationnel des ferments lactiques

P. Loubière, V. Laroute, P. Le Bourgeois,
ML Daveran-Mingot, M. Cocaign-Bousquet

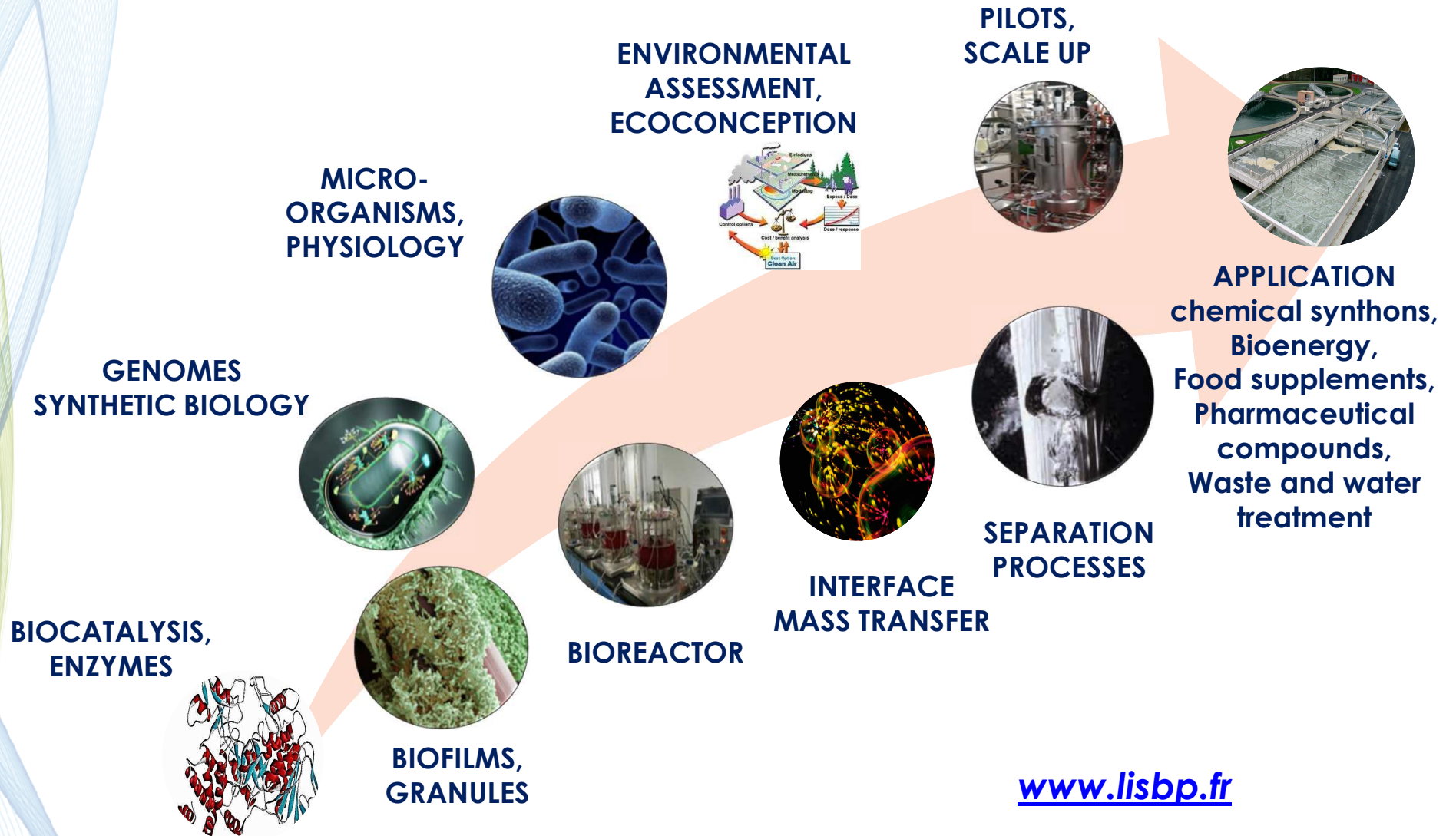
www.lisbp.fr

Toulouse, France



LISBP: a cascade of competences in biotechnology

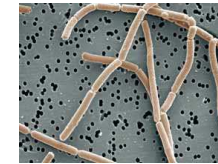
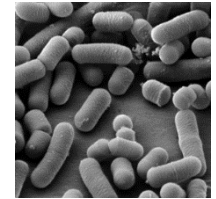
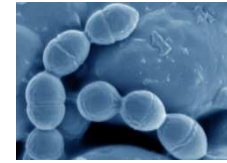
5 scientific poles, 4 platforms, 350 people



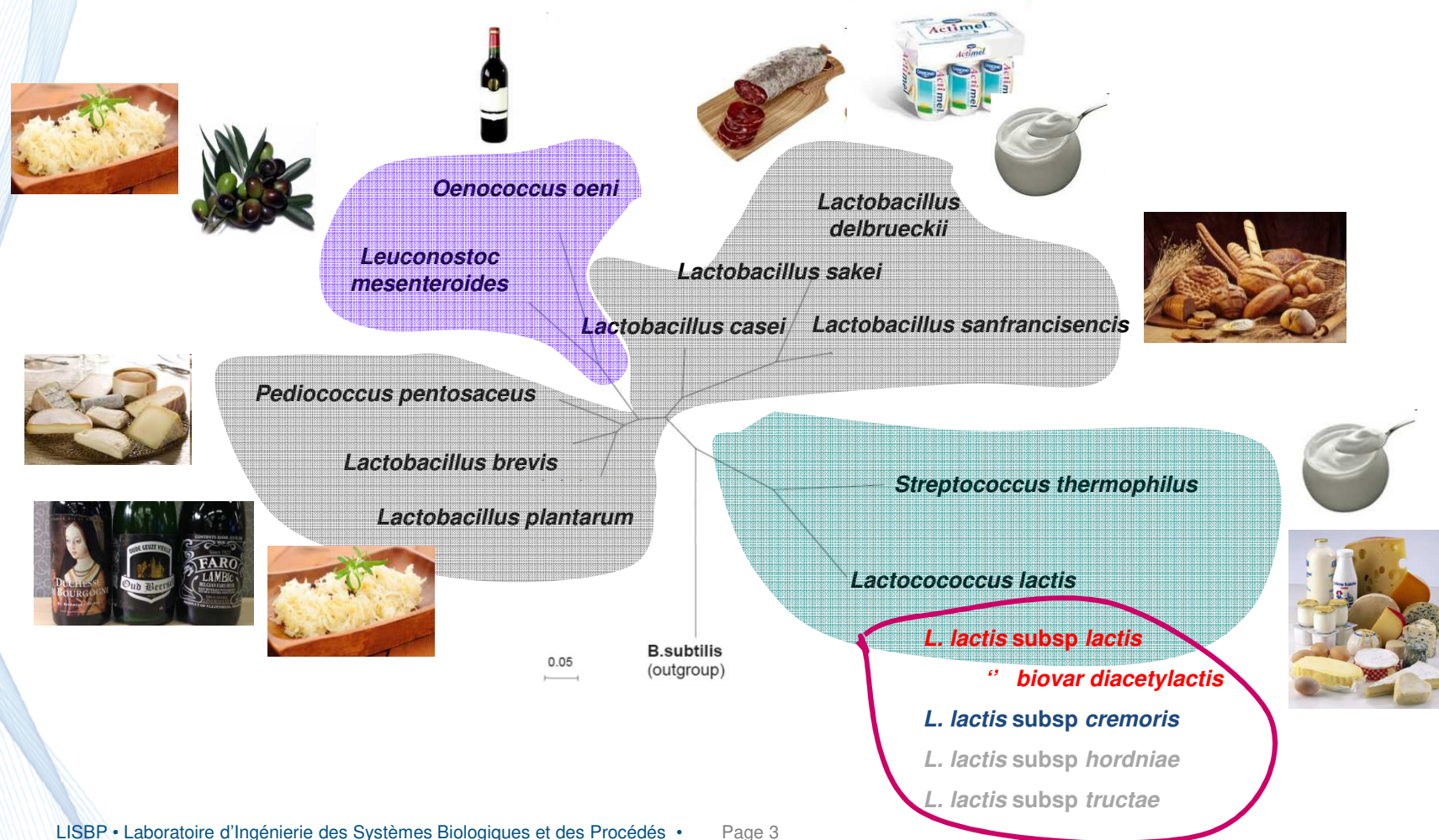
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Lactic Acid Bacteria

Gram-positive, non-spore forming cocci and rods
 Anaerobic aerotolerant, catalase negative
 GRAS (Generally Recognized As Safe) organisms

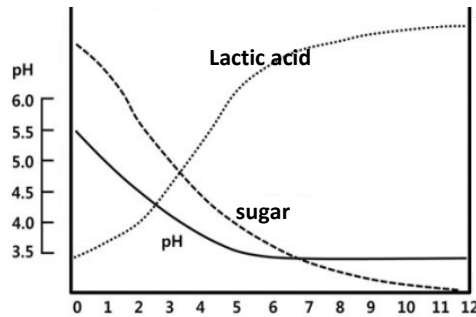


Large number of species for a variety of fermented foods

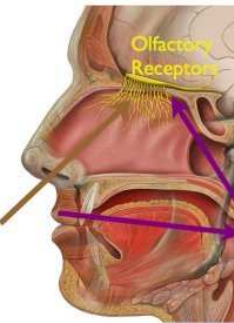


Lactic acid bacteria and food fermentation

Acidification (Lactic acid production)



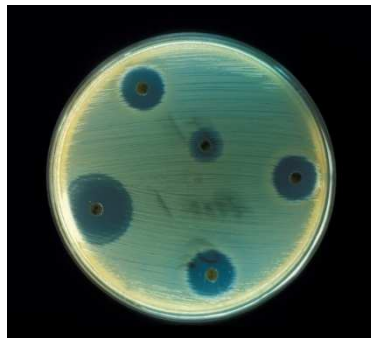
Flavor



Direct or orthonasal way

Olfactory Pathways

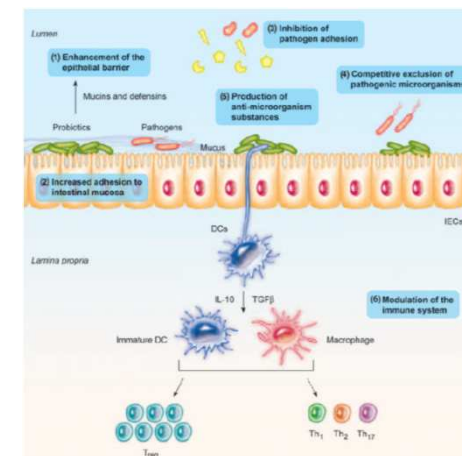
Antimicrobial compounds



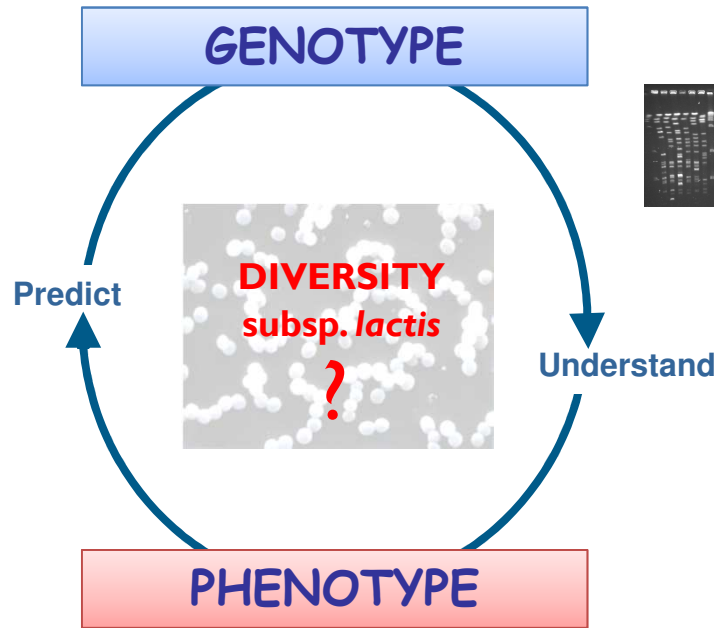
Texture



Nutraceutical production, Probiotic bacteria



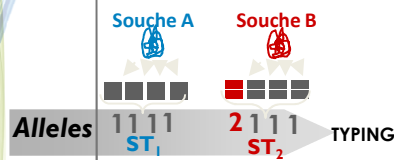
Diversity and functional characterization of starters: an integrated approach



Genetics

Genes content and genes variation

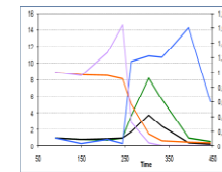
CGH (Comparative Genomic hybridation)
MLST (Multilocus sequence typing)



Genomics

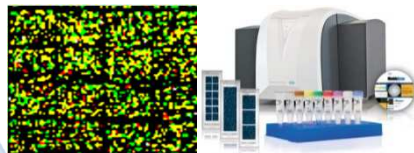
Genome sequencing **NGS**
Genome pattern analysis
PFGE (Pulsed Field Gel Electrophoresis)

Physiology



Growth, metabolites and acidification kinetics
Enzymatic analysis
Microbial energetics

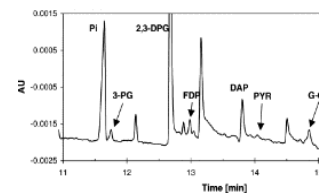
Transcriptomics



Proteomics



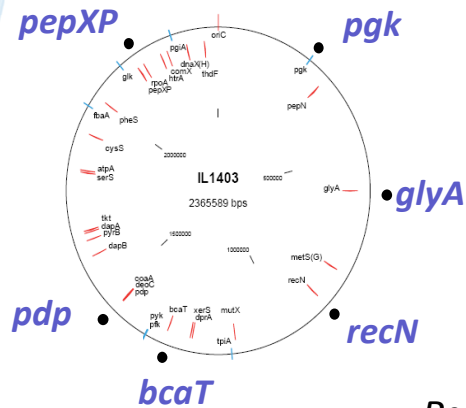
Metabolomics



Genetic diversity of *Lactococcus lactis* subsp *lactis* strains

MLST (Multi Locus Sequence Typing)

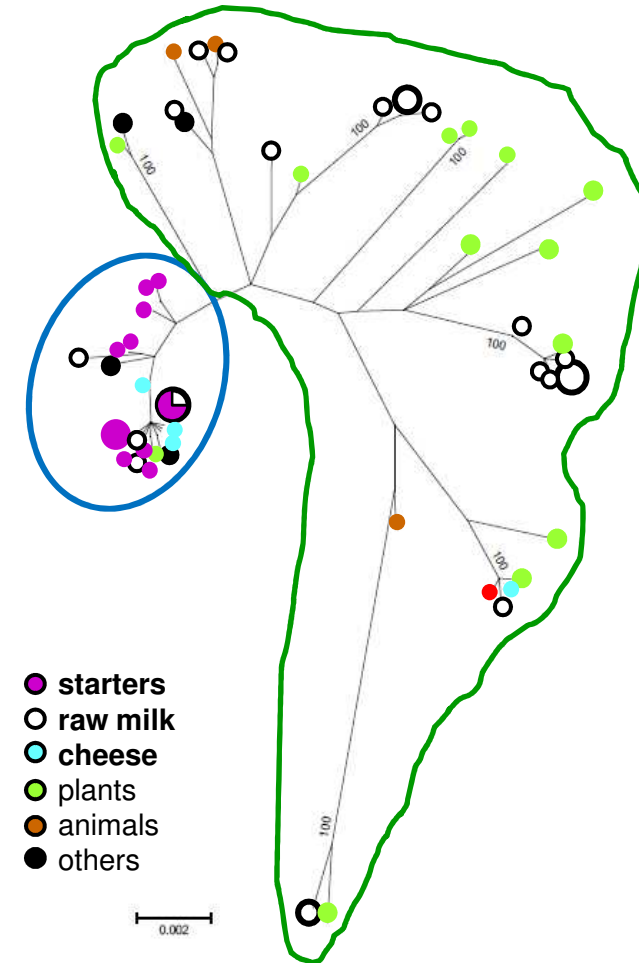
6 loci



154 strains -> 70 ST
L. lactis subsp. *lactis*

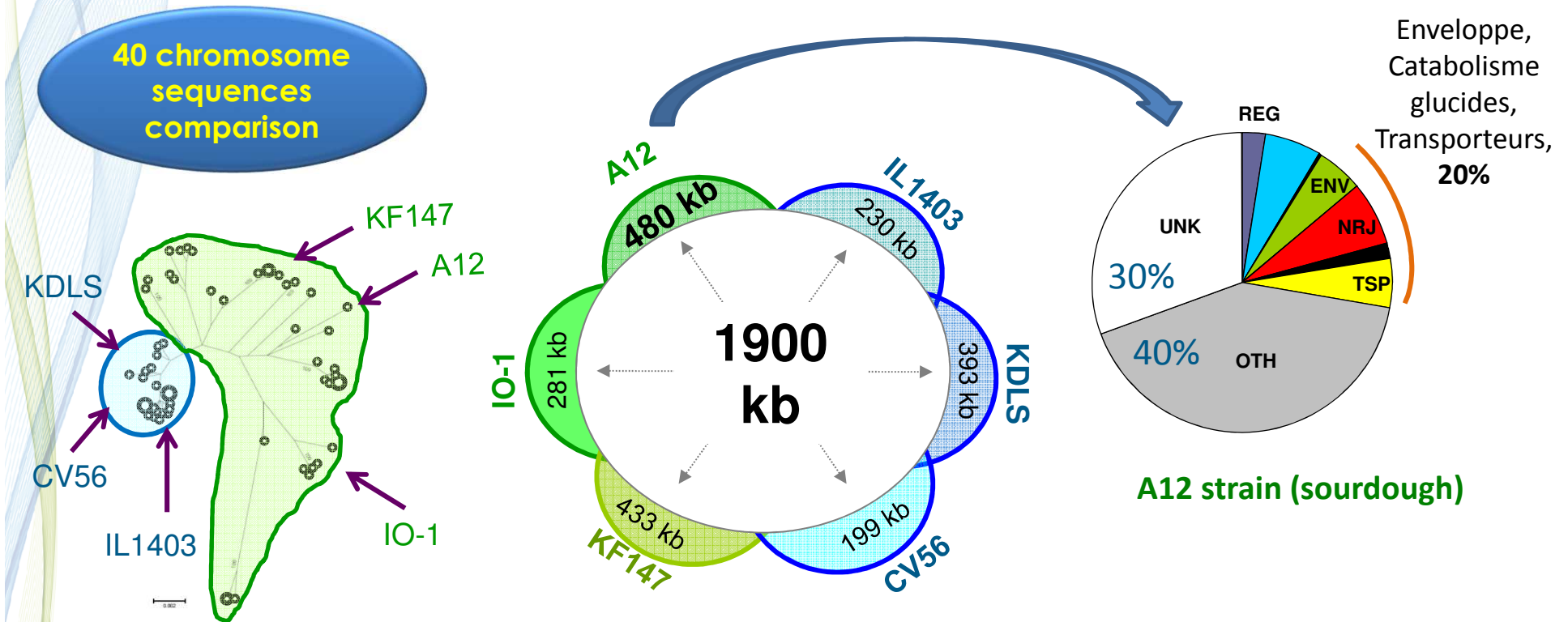
Passerini et al., Plos One 2010

- **Environmental strains**
high genetic diversity
plants, animals, raw milk
- **Domesticated strains**
low genetic diversity
industrial processes (starters, cheeses)



ML (T92+G+I) – bootstrap 500

Genomic diversity of *L. lactis* subsp *lactis* strains

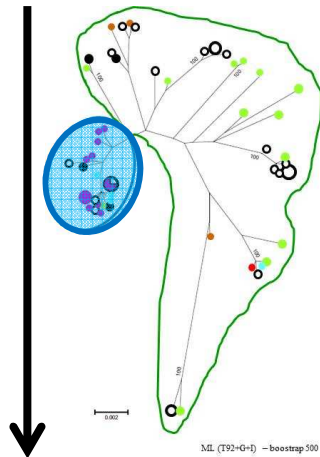


- « Strain-specific » genomic content (200 to 480 kb)
- A12 strain = 20% of genes related to sugar transport /catabolism
Efficient metabolism of plant sugars (*i.e.*, raffinose) → adaptation to sourdough

(Passerini et al., AEM 2013)

Functional diversity of *L. lactis* subsp *lactis* strains

Six “domesticated” strains



Cultivated as a model Cheese

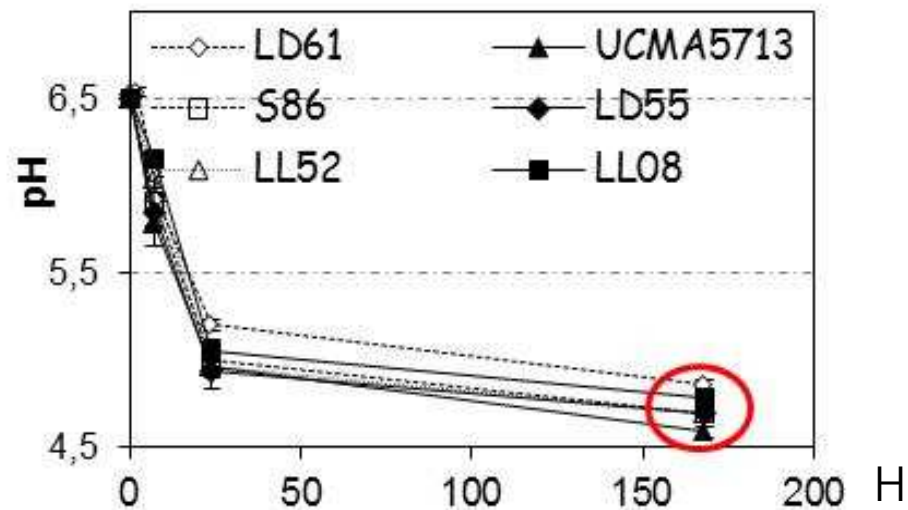


Ultra-filtered cheese

Six different phenotypes:
pH of the cheese at 7 days varies from 4.1 to 4.6

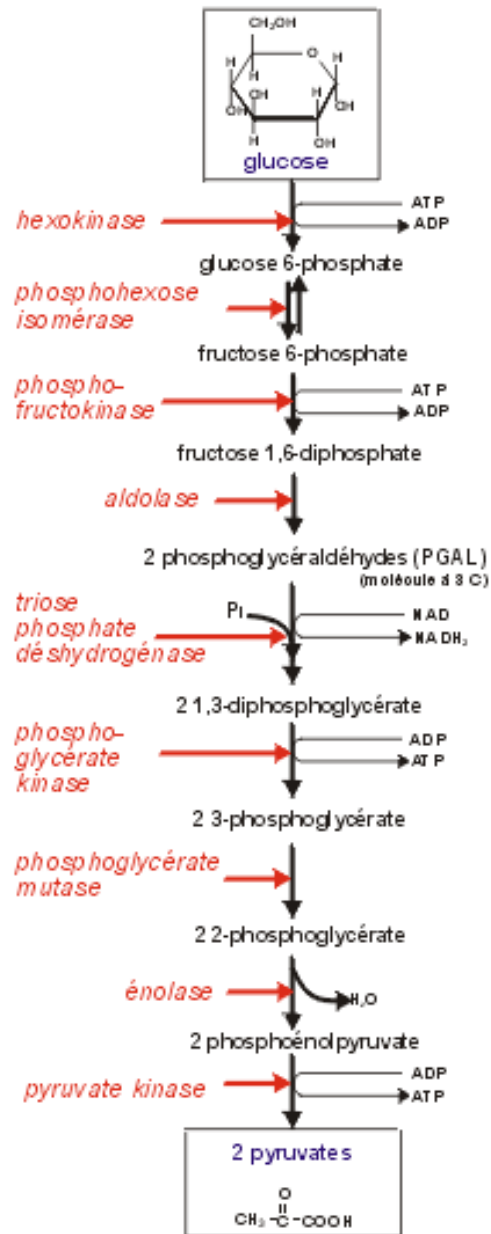


Significant differences in properties (texture, flavor) and bacterial viability



Tan-a-Ram et al., AEM 2011

Metabolic and energetic analysis of *L. lactis* subsp *lactis* strains



Metabolic analysis:

- **enzyme activities** all along the culture
- In vivo enzyme characterization : effect of external factors (pH, T°C, cofactors...)
 - Kinetic analysis of these factors
 - In vivo **carbon flux** measurement

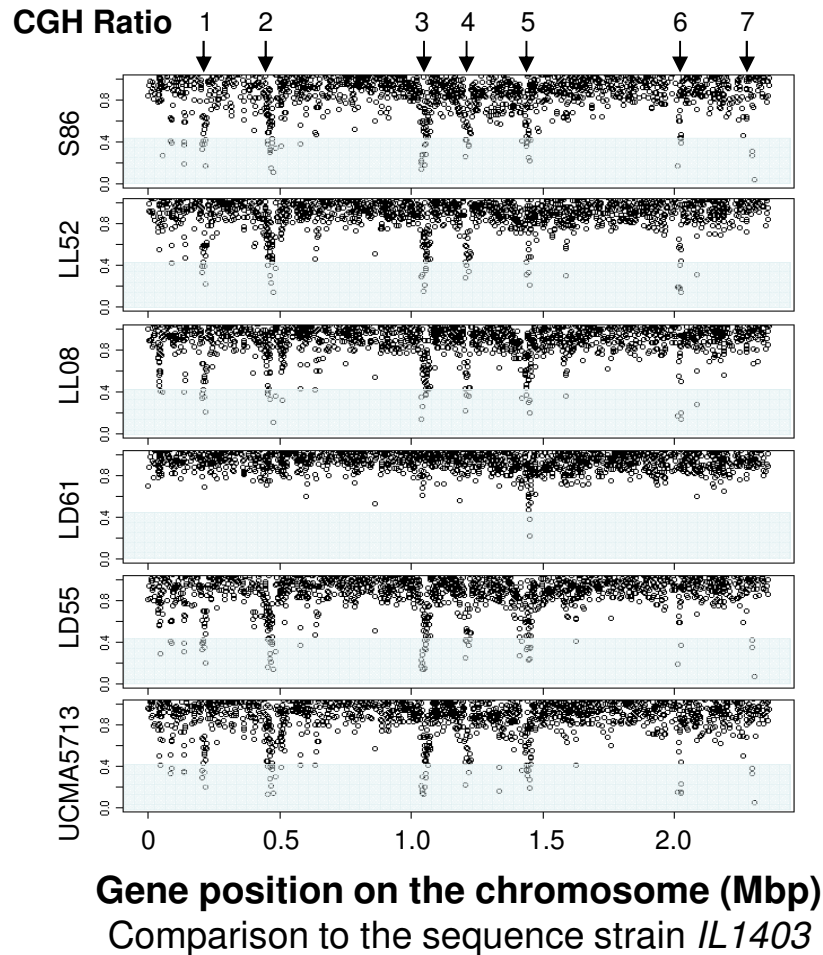


Modelisation of glycolysis in the culture, during growth, at growth arrest, and at post-acidification arrest

= **identification of limiting enzymes**, inhibited by low pH (PFK, PK)

Functional diversity of *L. lactis* subsp *lactis* strains

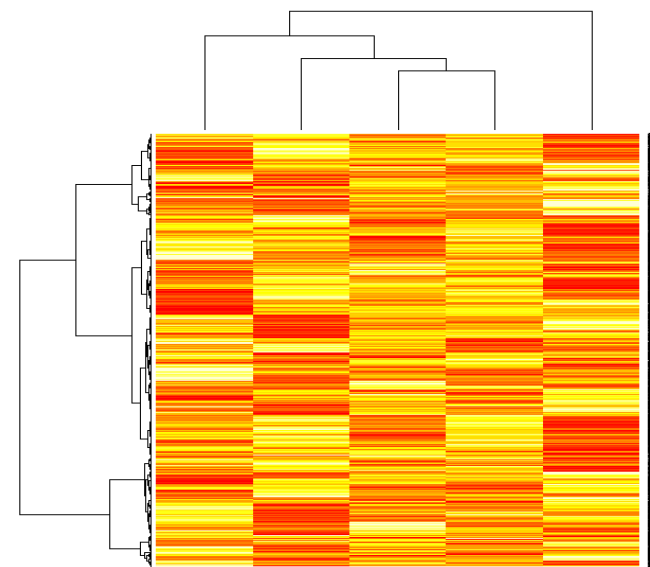
Core genome of 1915 genes
between the 6 strains.
Few genetic differences by CGH



Tan-a-Ram et al., AEM 2011

Dynamic transcriptome of *L. lactis*

Wide expression changes in the core genome,
between 250 and 600 genes differentially
expressed vs LD61



Differential expression in:
Response to acidic or oxidative stress,
Carbon limitation,
Carriers,
Some metabolic features (cit, arg, glu, ATPase...)

Functional analysis of *L. lactis* subsp *lactis* strain

Kinetic transcriptome of *L. lactis* LD61 in UF-cheese

Genes involved	Positive	Negative	
Growth rate response <i>rpl B,M,Q,U,V rpmGB,J rpsA,B,C,J,K,L,N,N2</i>			} Growth arrest and stress
General stress response <i>clpE, clpP, groES, hrcA-grpE-dnaK regulon, ytgH</i>			
Amino acid starvation response <i>oppA,B,C,D,F optC,D,S pep O,N,C,XP, pepXP,O,N,C</i>			
Acidic response <i>atp G,D, atpA,B,E,F,H arcC1 arcD1,D2</i>			Moderate acidic stress
Carbon starvation response <i>lacC,Z galM,K,T msmK rbsB celB ptbA ptcA,B,C,N,D scrK xylA bglA..</i>			Carbon limitation



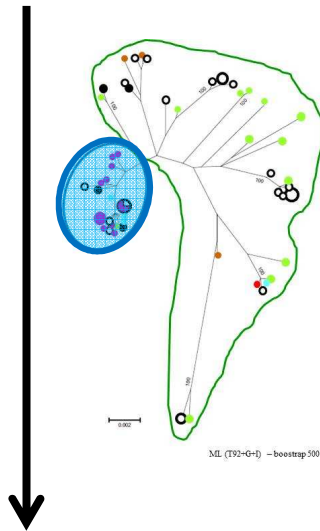
**Low diffusion
of lactose
inside the cheese**

Laroute et al, AEM, 2011

Functional diversity of *L. lactis* ssp *lactis* strains

Deep phenotype investigation of *L. lactis*

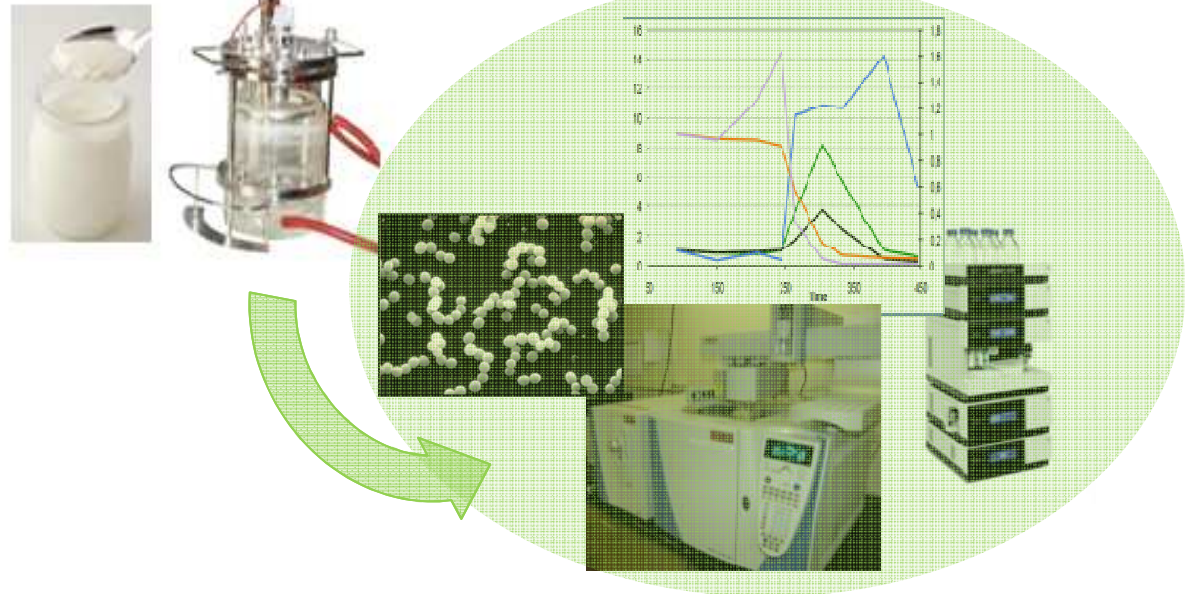
Nine
“domesticated” strains



In a model dairy product
(fermented milk)



Fermented milks



Physiological descriptors and metabolite concentrations
in dynamic and at 14 Days

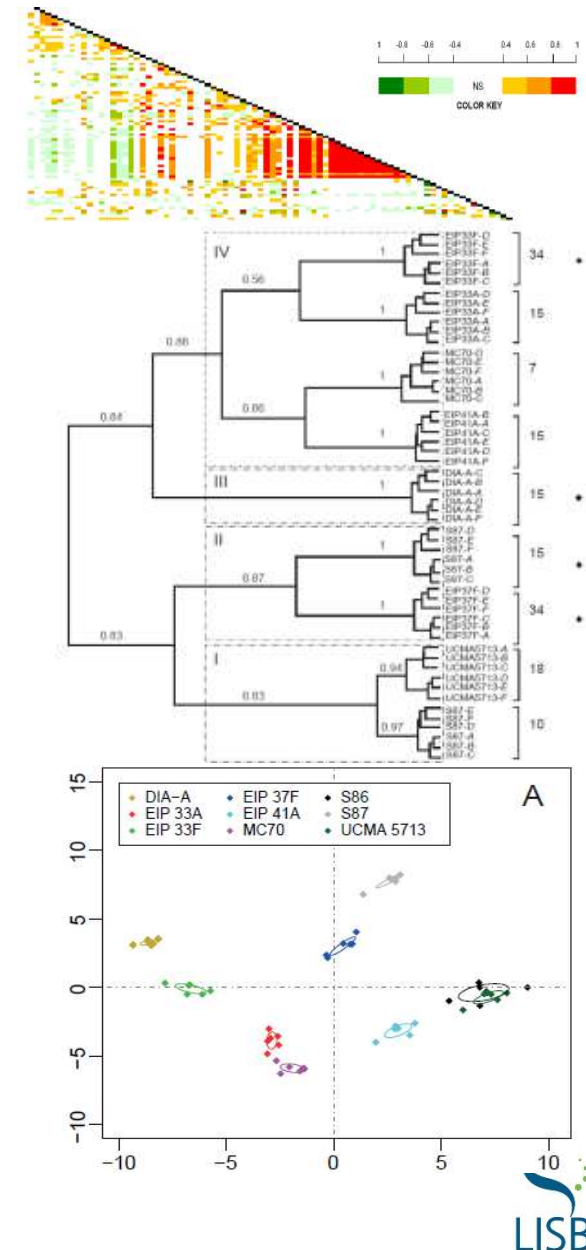
Dhaisne et al. , AEM, 2013

Functional diversity of *L. lactis* ssp *lactis* strains

82 variables
selected as important
features in dynamic
during 24h and at 14 Days

- Sugars,
- Organic acids (lactate, acetate, formate, acetaldehyde),
- Amino Acids ,
- 36 VOCs: Volatile Organic Compounds,
- Growth rate, acidity & redox activity

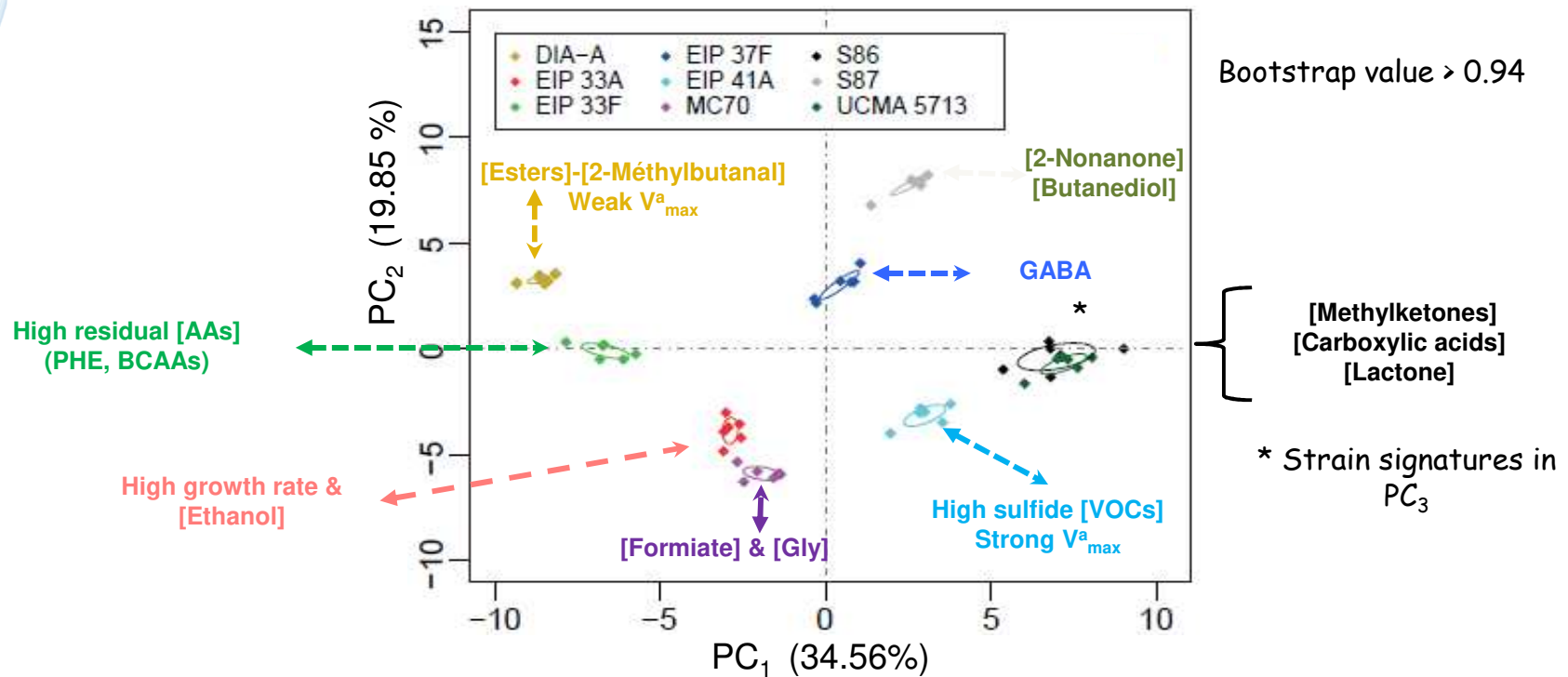
Statistical analysis
Unsupervised: PCA, HAC
Supervised: PLS-DA,
Random Forest



Functional diversity of *L. lactis ssp lactis* strains

Nine strains = nine signatures

↳ Accurate strain identification



Principal Component Analysis of the 82 phenotypic variables. Ellipses show strain category at a 95% confidence level.

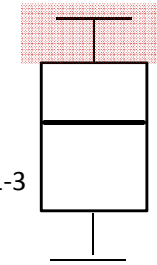
Functional diversity of *L. lactis ssp lactis* strains

82 variables

Earn time & money: variable selection

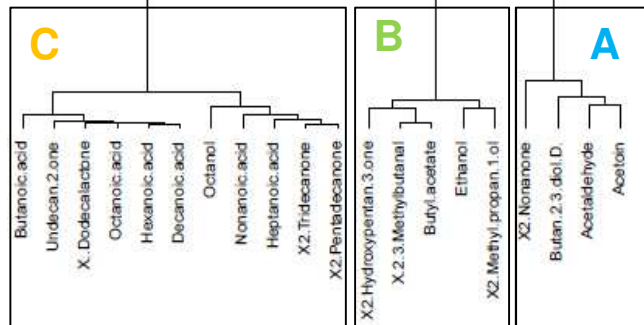
20 variables

Variable selection : 25% of the most contributive variables on PC₁₋₃



Variable clustering

20 variables = 20 VOCs

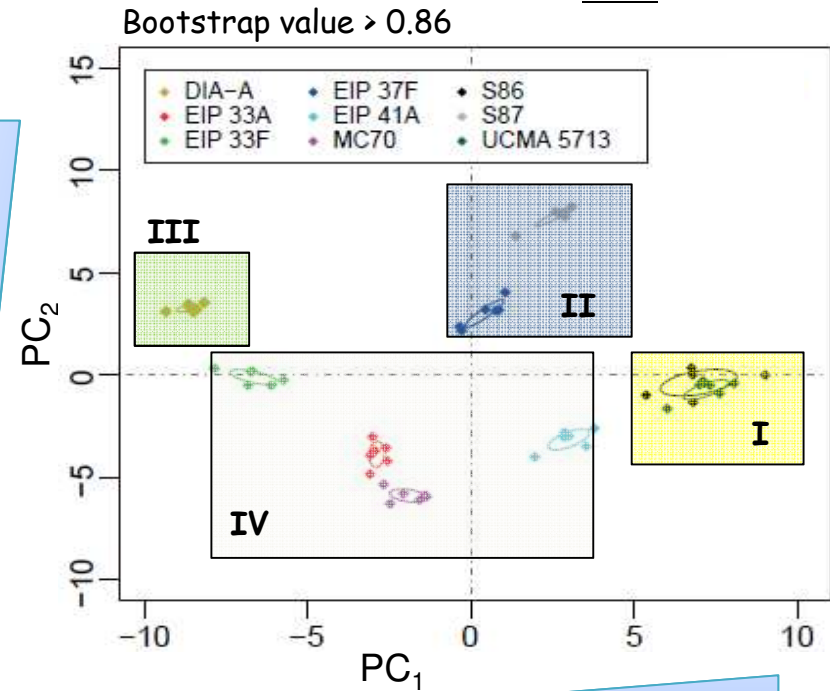


Lypolysis

Proteolysis

Glycolysis

Glycolysis



Lypolysis

Proteolysis

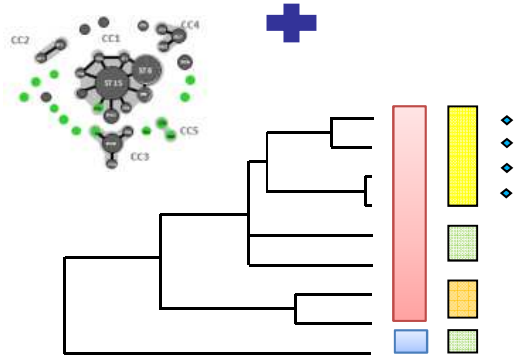
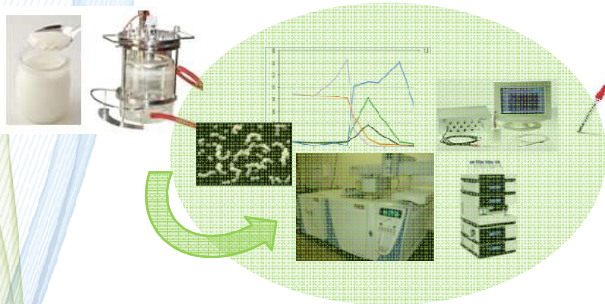
Four robust strain clusters

Functional diversity of *L. lactis ssp lactis* strains

Integration.... towards prediction

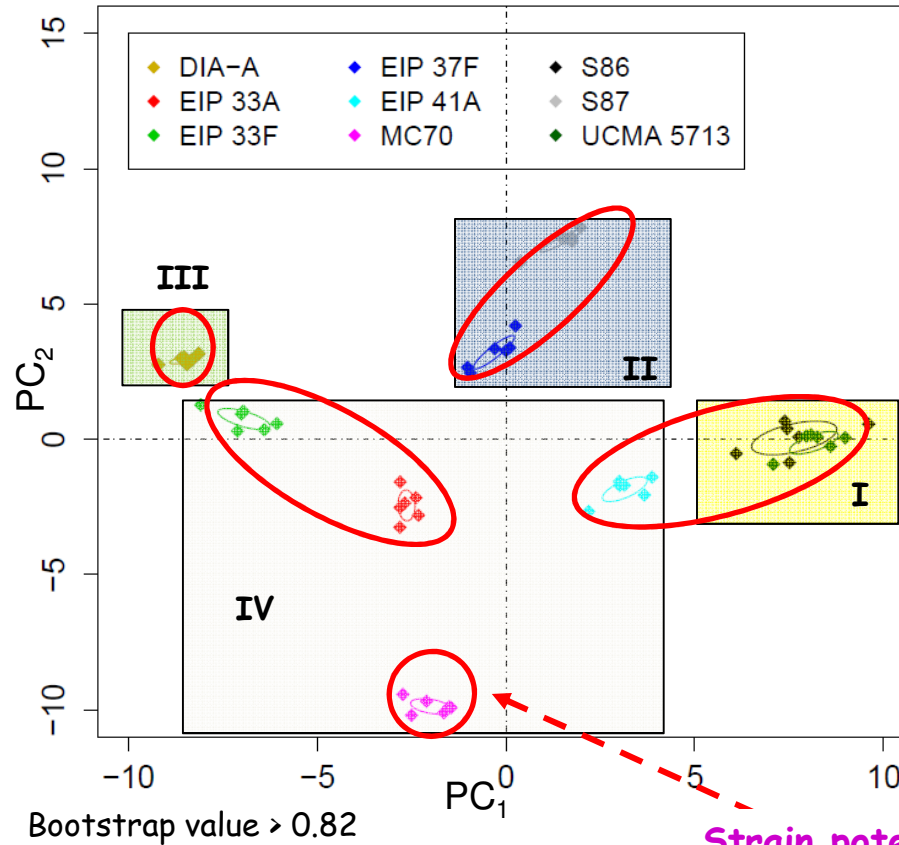
Dhalsne et al.
AEM 2013

Phenotype



Genotype

Inclusion of genotypic diversity led to 5 rather than 4 clusters



Strain potentialities encoded in the genotype

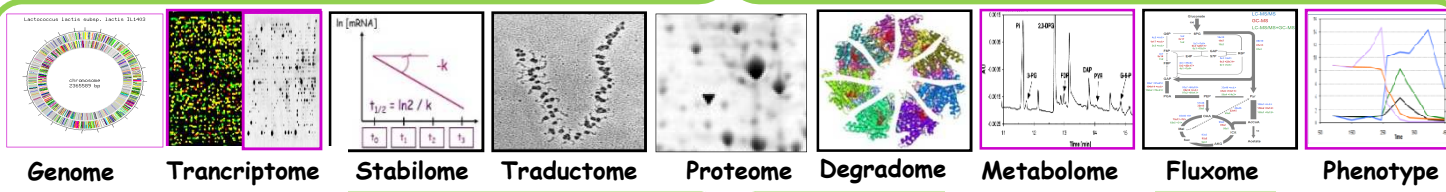
Fermented Food product



Natural strain diversity
and/or Starter addition

functional
responses

Data



Tools

INTEGRATION

Real understanding of bacterial potentialities to
strain/starter selection & phenotype prediction

Acknowledgements



Génie du métabolisme des procaryotes

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Pascal Le Bourgeois



Génétique des bactéries lactiques

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Maéva Guéllérin
Michèle Coddeville
Paul Ritzenthaler



Catherine Fontagné-Faucher
Valérie Gabriel
Hervé Robert



Hélène Tormo
Christel Couderc

Methodological & analytical support



Transcriptomics
Véronique Le Berre
Lidwine Trouilh



Metabolomics
JC. Portais
S Massou



Sébastien Déjean
Philippe Besse



Valentin Loux



Gilles De Revel

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